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Synthesis and Antiviral Effects of 2-Heteroaryl Substituted Adenosine and 8-Heteroaryl Substituted Guanosine Derivatives

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Abstract—2-(2"- and 3"-Thienyl)adenosine and the corresponding furyl derivatives were prepared through Pd(0)-catalyzed coupling of 2',3',5'-tri-O-(t-butyldimethylsilyl)-2-iodoadenosine with the appropriate tributyltin derivatives followed by deprotection. Preparation of the 8-(2"- and 3"-thienyl)guanosines and 8-(2"- and 3"-furyl)guanosines followed a similar route. Antiviral properties of these compounds and the related 2,6-diaminopurine ribofuranosides were of no pharmacological interest.

Introduction

We have previously found that 5-heteroaryl-substituted 2'-deoxyuridines show interesting antiviral properties. The triphosphates of some of them are potent inhibitors of HIV reverse transcriptase (RT).¹⁻² However, in a cell-culture assay the corresponding unphosphorylated nucleoside analogs were not active against HIV^{2,3} and the analogs with 3',5'-di-p-toluoyl protecting groups were inhibitory but with poor selectivity.³ The 5-heteroaryl-substituted 2'-deoxyuridines also have antiherpes activities.^{1,4} In a recent paper it was found that oligonucleotides containing 5-heteroaryl-substituted 2'-deoxyuridines gave enhanced thermal stability to complementary RNA relative to thymidine.⁵

Various purine nucleosides with modified sugars have shown good antiviral properties and 2',3'-dideoxyinosine has been approved for the treatment of acquired immunodeficiency syndrome (AIDS).⁶ 9-(2-Hydroxyethoxymethyl)guanine (ACV),⁷ 2',3'-dideoxy-2'-fluoropurine nucleosides⁸⁻¹⁰ and also 3'-fluoro-substituted purine nucleosides have been prepared as potent antiviral agents (for review see Ref. 11). 2-Substituted 2'-deoxyadenosine compounds have anti-metabolic properties. The 2-chloro analog is a potent antileukemic agent¹² and other 2-halo-adenine nucleosides have also been investigated.¹³

However, when we started our present work no systematic structure—activity study of purine nucleosides, having heterocyclic rings in the 2-, 6- and/or 8-positions, had been carried out. Recently, however, two compounds, 8-(2"-thienyl)- and 8-phenyl-adenosine, have been prepared and designed as possible drugs for treatment against sleeping sickness. ¹⁴ During recent years, an increased interest in the study of antiviral activity of modified purine nucleosides, substituted in the purine ring, has arisen.

Direct C-8-lithiation with lithium disopropylamide of sugar protected adenosine, inosine and guanosine has been used for the preparation of a number of 8-substituted derivatives. 15-17 Purine nucleosides alkylated in the 2- and 8-positions have been prepared by the palladium-catalyzed coupling of 2- and 8-bromoadenosine and 8-bromoguanosine derivatives with trialkylaluminum derivatives. 18 Alternatively, tetraalkyltin derivatives have been used in a Pd-catalyzed coupling of 2-, 6- and 8-halogenated purine nucleosides in order to prepare the corresponding alkyl derivatives. 19,20 Vinylation and allylation in the 8-position has also been achieved by Pd(0)-catalyzed coupling between 8-iodo-derivatives of t-butyl-dimethylsilyl protected adenosine, 2'-deoxyadenosine and 2',3'dideoxyadenosine and vinyltributyltin or allyltributyltin.²¹ 2-Alkynyladenosines were prepared in 1985 by Pd-catalyzed cross-coupling of 2-iodoadenosine and terminal acetylenes.²² The coupling of protected 2iodoadenosine with tributylstannyl acetone was utilized for the synthesis of 2-acetonyladenosine.²³ In a recent paper, the synthesis of 8-alkynyl-2'-deoxyadenosine analogs through Pd-catalyzed cross-coupling of 8bromo-2'-deoxyadenosine with terminal alkynes in the presence of copper(I) iodide in N,N-dimethylformamide was described. These compounds were converted by catalytic hydrogenation to the corresponding 8-alkenyland 8-alkyl derivatives, and the antiviral activities were studied.24 Purine derivatives, such as 6-alkoxypurine 2',3'-dideoxynucleosides have recently been found to inhibit the cytopathic effect of the HIV virus.25

In this paper, we describe the synthesis and the antiviral properties of 2-thienyl- and furyl-substituted adenosines, and of 8-thienyl- and furyl-substituted guanosines. Antiviral properties of the related 2,6-diaminopurine ribofuranosides 1a-d are also reported. Their synthetic route is described elsewhere.²⁶

$$H_{2} \stackrel{\text{MH}_{2}}{\longrightarrow} \stackrel{\text{MH}_{2}}{\longrightarrow} \stackrel{\text{M}}{\longrightarrow} \stackrel{\text{$$

Chemistry

a - d

2-(2"-and 3"-Thienyl)adenosine (5a, b) and corresponding furyl derivatives (5c, d) were prepared through Pd(0)-catalyzed coupling of 2',3',5'-tri-O-(tbutyldimethylsilyl)-2-iodoadenosine (3) with the appropriate tributyltin derivatives followed by deprotection. Compound 2 was prepared from guanosine by acetylation followed by chlorination to 9-(2',3',5'tri-O-a c e tyl-β-D-ribofuranosyl)-2-amino-6-chloropurine according to Robins and Uznanski.27 This compound was then iodinated in the 2-position by heating with pentyl nitrite and diiodomethane to 9-(2',3',5'-tri-Oacetyl-β-D-ribofuranosyl)-2-iodo-6-chloropurine, 28 which was treated with ethanolic ammonia at 0 °C yielding 2iodoadenosine (2).29 The hydroxyl groups of 2 were protected through reaction with t-butyldimethylsilyl chloride to give 3.30 The Pd(0)-catalyzed coupling of 3 with five equivalents of 2- or 3-tributylstannylthiophene or 2- or 3-tributylstannylfuran was carried out using PdCl₂(PPh₃)₂ in refluxing tetrahydrofuran^{31,32} and the reaction times were about 50 h. This strategy gave the

t-butyldimethylsilyl protected compounds 4a-d in 82-95% yields. Deprotection of the nucleosides was carried out by treatment with 0.1 M methanolic hydrogen chloride according to Olgivie et al.³⁰ yielding 5a-d in 62-72% yield. The 2"-thienyl derivative 5a has previously been prepared by reaction of 2-cyanothiophene with the riboside of 5-amino-4-cyanoimidazole (AICN-riboside) in methanolic ammonia;^{33,34} however, no yield was given.

Preparation of the 8-(2"- and 3"-thienyl)guanosines and 8-(2"- and 3"-furyl)guanosines (7a-d) also started from guanosine, which was brominated to 8-bromoguanosine (6). As described for the adenosine derivatives, the sugar was protected with t-butyldimethylsilyl groups. The Pd(0)-catalyzed coupling with 2- or 3-tributyl-stannylthiophene or 2- or 3-tributylstannylfuran could be carried out successfully. However, upon attempted deprotection with 0.1 M methanolic hydrogen chloride, the glycosidic bond was cleaved. According to Ratsep et al. The presence of strong electron-withdrawing groups in the 8-position makes the glycosidic

i. t-BuMe₂SiCl, Imidazole, DMF
 ii. YSn(Bu)₃, PdCl₂(PPh₃)₂, THF, Δ
 iii. 0.1 M HCVMeOH

bond very acid labile. Although thienyl and furyl groups can be both electron-withdrawing or electron-donating, we hardly expected such a strong effect on the glycosidic bond. Finally, we used a modification of the method described by Mamos et al. 19 consisting of in situ protection of 5 by refluxing with hexamethyl-disilazane, followed by Pd(0)-catalyzed coupling of the crude product with 2- or 3-tributylstannylthiophene or 2- or 3-tributylstannylfuran. 32

Compounds 7a-d were obtained in 52-58% yield after deprotection with potassium carbonate in methanol at room temperature. Further purification of 7a-d was achieved by HPLC chromatography on a reversed-phase column using acetonitrile:water (20:80) as eluent.

Inhibition of Viral Replication in Cell Culture Assays

Compounds 1a-d, 2, 4a-d, 5a-d and 7a-d were tested in cell culture assays for their inhibition of multiplication of human immunodeficiency virus 1 (HIV-1), herpes simplex virus-1 (HSV-1), human cytomegalovirus (HCMV) and influenza virus A. In addition, the effect on the growth of uninfected cells was determined. The activities of the compounds were studied at 1, 10 and $100 \ \mu g \ mL^{-1}$ concentrations.

All compounds were inactive against HSV-1. Two compounds, 1b and 1d had weak activities against influenza A, showing 50% inhibition at the highest concentration, 100 μ g mL⁻¹. In the HIV-1 assay, only compound 2 showed some activity and was retested at lower concentrations (0.1, 0.01 and 0.001 mg mL⁻¹). The 50% inhibitory concentration, ED₅₀, was found to be 0.5 μ g mL⁻¹. However, at 100 μ g mL⁻¹ the compound completely inhibited cellular proliferation. Compounds 5a, 1b and 1d exhibited some activity against HCMV, both in an ELISA and cytopathogenic (CPE) assay. In the ELISA assay, the ED₅₀s were 12, 14 and 20 μ g

mL⁻¹, respectively. In the CPE assay, **5a** showed a > 75% inhibition at 100 μ g mL⁻¹. However, in the virus free assay, cellular proliferation was also inhibited (95%) at 100 μ g mL⁻¹ by this compound. For compounds **1b** and **1d**, no definite antiviral cytopathogenic effect could be seen at 100 μ g mL⁻¹, due to interfering cellular toxicity.

In the cell growth assay, all compounds had some inhibitory activity at the highest concentration, 100 μ g mL⁻¹. Compounds **4a**, **4b**, **4c**, **5a**, **1a**, **1b** and **1d** showed about 95% inhibition each, while compounds **2**, **4d**, **5b**, **5c**, **5d** and **1c** about 50% inhibition and compounds **7a**–**d** about 30% inhibition.

Experimental

Chemistry

The ¹H NMR spectra were recorded on a Varian XL-300 spectrometer. The mass spectra were recorded on a JEOL JMS-SX 102 spectrometer. Melting points are uncorrected. Flash column chromatography was carried out using Merck silica gel 60. Tetrahydrofuran (THF) was dried by refluxing and distillation over sodium wire. N,N-Dimethylformamide (DMF) was dried over 4 Å molecular sieves. Pyridine was distilled and stored over 4 Å molecular sieves. All other solvents were distilled prior to use.

Preparation of 9-[2',3',5'-tri-O-(t-butyldimethylsilyl)- β -D-ribofuranosyl]-6-amino-2-iodopurine (3). To a mixture of 0.96 g (6.4 mmol) of t-butyldimethylsilylchloride and 0.89 g (13 mmol) of imidazole in 4.0 mL of anhydrous DMF, 0.50 g (1.3 mmol) of 9-(β -D-ribofuranosyl)-6-amino-2-iodopurine (2) was added. The resulting solution was stirred for 28 h at room temperature. When no starting material remained (checked by thin-layer chromatography (TLC), toluene:ethylacetate 4:1), the mixture was poured into an ethyl acetate/water solution. The organic layer was separated, dried, and evaporated

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to dryness. The residue was chromatographed using toluene:ethyl acetate (4:1) as eluent, to give 0.65 g (68%) of the title compound as a white solid. H NMR (CDCl₃) showed the same δ values as those given in the literature.³⁷

General procedure for preparation of 2-(2"-heteroaryl)-9-[2',3',5'-tri-O-(t-butyldimethylsilyl)- β -D-ribofuranosyl]-adenine. A mixture of 3, PdCl₂(PPh₃)₂ and tributyl-stannylheteroaryl³² in anhydrous THF was heated with stirring under nitrogen at 90 °C. The reaction was followed by TLC using dichloromethane:methanol (9:1) as eluent. When the starting material was consumed, the reaction mixture was allowed to reach room temperature and was evaporated to dryness. The residue was column chromatographed using toluene:ethyl acetate (4:1) as eluent. Further purification was achieved by HPLC using a nucleosil column (500 × 10 mm) and toluene:methanol (7:3) as eluent to give the desired compound.

2-(2"-Thienyl)-9-[2',3',5'-tri-O-(t-butyldimethylsilyl)-β-D-ribofuranosyl]adenine (4a). This compound was obtained from 250 mg (0.34 mmol) of 3, 14 mg (0.02 mmol) of PdCl₂(PPh₃)₂ and 634 mg (1.70 mmol) of 2-tributylstannylthiophene³² in 2.5 mL of anhydrous THF. After 48 h, work up and purification by HPLC the yield was 193 mg (82%); mp 235–237 °C. ¹H NMR (CDCl₃): δ 8.14 (s, 1H, H8), 7.90 (dd, 1H, H3", J = 3.7, 1.2 Hz), 7.38 (dd, 1H, H5", J = 5.0, 1.2 Hz), 7.10 (dd, 1H, H4", J = 5.0, 3.7 Hz), 6.00 (d, 1H), 5.54 (s, NH₂), 4.81 (t, 1H, H2'), 4.36 (t, 1H, H3'), 4.10 (m, 1H, H5'), 3.84 (d, 1H, H4'), 0.97–0.82 (m, 27H, Si-t-Bu), 0.16–0.10 (m, 18H, SiMe₃). Peak matching for (M⁺ + H), calcd for C₃₂H₅₈O₄N₅Si₃S: 692.3517; found: 692.3526.

2-(3"-Thienyl)-9-[2',3',5'-tri-O-(t-butyldimethylsilyl)-β-D-ribofuranosyl]adenine (4b). This compound was obtained from 250 mg (0.34 mmol) of 3, 14 mg (0.02 mmol) of PdCl₂(PPh₃)₂ and 634 mg (1.70 mmol) of 2-tri-butylstannylthiophene³² in 2.5 mL of anhydrous THF. After 48 h, work up and purification by HPLC the yield was 209 mg (89%); mp 190–191 °C. ¹H NMR (CDCl₃): δ 8.18 (s, 1H, H8), 8.16 (dd, 1H, H2", J = 2.9, 1.1 Hz), 7.86 (dd, 1H, H4", J = 5.0, 1.1 Hz), 7.32 (dd, 1H, H5", J = 5.0, 2.9 Hz), 6.06 (d, 1H, H1'), 5.48 (s, NH₂), 4.73 (t, 1H, H2'), 4.35 (t, 1H, H3'), 4.13 (m, 1H, H5'), 3.85 (d, 1H, H4'), 0.97–0.82 (m, 27H, Si-t-Bu), 0.16–0.10 (m, 18H, SiMe₃). Peak matching for (M* + H), calcd for C₃₂H₅₈O₄N₅Si₃S: M_r 692.3517; found: 692.3526.

2-(2"-Furyl)-9-[2',3',5'-tri-O-(t-butyldimethylsilyl)-β-D-ribofuranosyl]adenine (4c). This compound was obtained from 250 mg (0.34 mmol) of 3, 14 mg (0.02 mmol) of PdCl₂(PPh₃)₂ and 606 mg (1.70 mmol) of 2-tributylstannylfuran³² in 2.5 mL of anhydrous THF. After 72 h, work up and purification by HPLC the yield was 216 mg (94%); mp 252-255 °C. ¹H NMR (CDCl₃): δ 8.20 (s, 1H, H8), 7.57 (dd, 1H, H5", J = 1.8, 0.9 Hz), 7.18 (dd, 1H, H3", J = 3.4, 0.9 Hz), 6.52 (dd, 1H, H4", J = 3.4, 1.8 Hz), 6.03 (d, 1H, H1'), 5.65 (s, NH₂), 4.69

(dd, 1H, H2'), 4.35 (dd, 1H, H3'), 4.12 (m, 1H, H5'), 3.82 (dd, 1H, H4'), 0.98–0.82 (m, 27H, Si-t-Bu), 0.17–0.10 (m, 18H, SiMe₃). Peak matching for (M⁺ + H), calcd for $C_{32}H_{58}O_5N_5Si_3$: 676.3746; found: 676.3746.

2-(3"-Furyl)-9-[2',3',5'-tri-O-(t-butyldimethylsilyl)-β-D-ribofuranosyl]adenine (4d). This compound was obtained from 250 mg (0.34 mmol) of 3, 14 mg (0.02 mmol) of PdCl₂(PPh₃)₂ and 606 mg (1.70 mmol) of 3-tributylstannylfuran³² in 2.5 mL of anhydrous THF. After 72 h, work up and purification by HPLC the yield was 218 mg (95%); mp 214–215 °C. ¹H NMR (CDCl₃): δ 8.18 (s, 1H, H8), 8.11 (dd, 1H, H2", J = 1.5, 1.0 Hz), 7.44 (dd, 1H, H5", J = 1.8, 1.5 Hz), 7.01 (dd, 1H, H4", J = 1.8, 1.0 Hz), 6.03 (d, 1H, H1'), 5.48 (s, NH₂), 4.64 (dd, 1H, H2'), 4.34 (dd, 1H, H3'), 4.08 (m, 1H, H5'), 3.82 (dd, 1H, H4'), 0.98–0.80 (m, 27H, Si-t-Bu), 0.17–0.10 (m, 18H, SiMe₃). Peak matching for (M* + H), calcd for C₃₂H₅₈O₅N₅Si₃: M_r 676.3746; found: (M + 1) 676.3739.

General procedure for the deprotection of compounds 4a-d. Compounds 4a-d were each dissolved in 0.1 M methanolic hydrochloric acid and stirred at room temperature for 24 h. The reaction was followed by TLC using dichloromethane:methanol (9:1) as eluent. The solvent was evaporated and the residue column chromatographed using a gradient of dichloromethane: methanol (9:1-7:3) as eluent. Further purification was achieved on HPLC using a nucleosil (500 × 10 mm) column and chloroform:methanol as eluent.

2-(2"-Thienyl)adenosine (5a). This compound was obtained from 120 mg (0.17 mmol) of 4a and 30 mL of 0.1 M methanolic hydrochloric acid in a yield of 38 mg (63%); mp 244–246 °C (lit. 250 °C). The proportions for the eluent were 92:8. H NMR (DMSO- d_6): δ 8.34 (s, 1H, H8), 7.83 (dd, 1H, H3", J = 3.6, 1.3 Hz), 7.63 (dd, 1H, H5", J = 5.1, 1.3 Hz), 7.40 (s, NH₂), 7.15 (dd, 1H, H4", J = 5.1, 3.6 Hz), 5.91 (d, 1H, H1'), 4.72 (dd, 1H, H2'), 4.22 (dd, 1H, H3'), 3.96 (dd, 1H, H4'), 3.64 (m, 1H, H5'). Peak matching (M⁺ + H), calcd for $C_{14}H_{16}O_4N_4S$: 350.0923; found: 350.0929.

2-(3"-Thienyl)adenosine (5b). This compound was obtained from 120 mg (0. 17 mmol) of 4b and 30 mL of 0.1 M methanolic hydrochloric acid in a yield of 43 mg (72%); mp 185–188 °C. The proportions for the eluent were 92:8. ¹H NMR (DMSO- d_6): δ 8.35 (s, 1H, H8), 8.17 (dd, 1H, H2", J = 3.1, 1.2 Hz), 7.78 (dd, 1H, H4", J = 5.1, 1.2 Hz), 7.59 (dd, 1H, H5", J = 5.1, 3.1 Hz), 7.34 (s, NH₂), 5.93 (d, 1H, H1'), 4.74 (t, 1H, H2'), 4.22 (dd, 1H, H3'), 3.96 (dd, 1H, H4'), 3.61 (m, 1H, H5'). Peak matching for (M* + H), calcd for $C_{14}H_{16}O_4N_5S$: 350.0923; found: 350.0926.

2-(2"-Furyl)adenosine (5c). This compound was obtained from 207 mg (0.31 mmol) of 4c and 100 mL of 0.1 M methanolic hydrochloric acid in a yield of 64 mg (62%); mp 137-139 °C (lit. 135-140 °C).³³ The pro-

portions for the eluent were 90:10. ¹H NMR (DMSO- d_6): δ 8.36 (s, 1H, H8), 7.81 (dd, 1H, H5", J = 1.8, 0.8 Hz), 7.43 (s, NH₂), 7.13 (dd, 1H, H3", J = 3.4, 0.8 Hz), 6.63 (dd, 1H, H4", J = 3.4, 1.8 Hz), 5.92 (d, 1H, H1'), 4.68 (dd, 1H, H2'), 4.19 (dd, 1H, H3'), 3.97 (dd, 1H, H4'), 3.20 (m, 1H, H5'). Peak matching for (M^+ + H), calcd for $C_{14}H_{16}O_5N_5$: 334.1151; found: 334.1155.

2-(3"-Furyl)adenosine (5d). This compound was obtained from 201 mg (0.30 mmol) of 4d and 100 mL of 0.1 M methanolic hydrochloric acid in a yield of 67 mg (67%); mp 154–155 °C. The proportions for the eluent were 90:10. ¹H NMR (DMSO- d_6): δ 8.32 (s, 1H, H8), 8.21 (dd, 1H, H2", J = 1.6, 0.9 Hz), 7.74 (dd, 1H, H5", J = 1.8, 1.6 Hz), 7.32 (s, NH₂), 7.00 (dd, 1H, H4", J = 1.8, 0.9 Hz), 5.91 (d, 1H, H1'), 4.72 (dd, 1H, H2'), 4.20 (t, 1H, H3'), 3.98 (dd, 1H, H4'), 3.68 (m, 1H, H5'). Peak matching for (M* + H), calcd for $C_{14}H_{16}O_5N_5$: 334.1151; found: 334.1142.

General procedure for the preparation of 8-(heteroaryl)guanosine. A suspension of 8-bromoguanosine (6), 1,1,1,3,3,3-hexamethyldisilazan (HMDS) and pyridine in the presence of a catalytic amount of ammonium sulfate was refluxed overnight under an atmosphere of nitrogen. After cooling to room temperature, the volatiles were removed in vacuo to give the trimethylsilylprotected nucleoside as a syrup, which was used in the next step without purification. The crude product was dissolved in anhydrous THF, PdCl₂(PPh₃)₂ and the tributylstannylheteroaryl were added, after which the suspension was heated at 90 °C under an atmosphere of nitrogen until no starting material remained (checked by RP C_{18} -TLC using acetone:water (1:1) as eluent). After cooling to room temperature, the solvent was removed in vacuo and the crude product was filtered through a silica gel 60 column using dichloromethane:methanol (9:1) as eluent. The trimethylsilyl groups were removed by treatment with potassium carbonate in methanol at room temperature for 1 h. The reaction mixture was evaporated to dryness and the crude product was purified directly by HPLC on a dynamax RP C_{18} column (500 × 10 mm) using acetonitrile:water as eluent.

8-(2"-Thienyl)guanosine (7a). This compound was obtained from 250 mg (0.69 mmol) of 6, 25.0 mL of HMDS, 2.5 mL of pyridine, 25 mg (0.03 mmol) of PdCl₂(PPh₃)₂ and 1.29 g (3.45 mmol) of 2-tributyl-stannylthiophene.³² After 25 h and HPLC purification (eluent 20:80) the yield was 146 mg (58%); mp 249 °C (dec). ¹H NMR (DMSO- d_6): δ 7.74 (dd, 1H, H5", J = 5.1, 1.0 Hz), 7.48 (dd, 1H, H3", J = 3.6, 1.0 Hz), 7.21 (dd, 1H, H4", J = 5.1, 3.6 Hz), 5.85 (d, 1H, H1'), 5.45 (s, NH₂), 5.09 (t, 1H, H2'), 4.12 (dd, 1H, H3'), 3.88 (dd, 1H, H4'), 3.58 (m, 1H, H5'). Peak matching for (M⁺ + H), calcd for C₁₄H₁₆O₅N₅S: 366.0872; found: 366.0870.

8-(3"-Thienyl)guanosine (7b). This compound was obtained from 250 mg (0.69 mmol) of 6, 25.0 mL of HMDS, 2.5 mL of pyridine, 25 mg (0.03 mmol) of

PdCl₂(PPh₃)₂ and 1.29 g (3.45 mmol) of 3-tributyl-stannylthiophene.³² After 25 h and HPLC purification (eluent 23:77) the yield was 131 mg (52%); mp 270 °C (dec). ¹H NMR (DMSO- d_6): δ 7.89 (dd, 1H, H2", J = 2.9, 1.2 Hz), 7.72 (dd, 1H, H5", J = 5.0, 2.9 Hz), 7.44 (dd, 1H, H4", J = 5.0, 1.2 Hz), 6.66 (s, NH₂), 5.76 (d, 1H, H1'), 5.00 (t, 1H, H2'), 4.10 (dd, 1H, H3'), 3.88 (dd, 1H, H4'), 3.67 (m, 1H, H5'). Peak matching for (M⁺ + H), calcd for C₁₄H₁₆O₅N₅S: 366.0872; found: 366.0867.

8-(2"-Furyl)guanosine (7c). This compound was obtained from 250 mg (0.69 mmol) of 6, 25.0 mL of HMDS, 2.5 mL of pyridine, 25 mg (0.03 mmol) of $PdCl_2(PPh_3)_2$ and 1.23 mg (3.45 mmol) of 2-tributylstannylfuran.³² After 40 h and HPLC purification (eluent 20:80) the yield was 128 mg (53%); mp 255 °C (dec). ¹H NMR (DMSO- d_6): δ 7.89 (dd, 1H, H5", J = 1.7, 0.8 Hz), 6.92 (dd, 1H, H3", J = 3.4, 0.8 Hz), 6.68 (dd, 1H, H4", J = 3.4, 1.7 Hz), 6.68 (s, NH₂), 5.94 (d, 1H, H1'), 5.02 (t, 1H, H2'), 4.13 (dd, 1H, H3'), 3.89 (dd, 1H, H4'), 3.67 (m, 1H, H5'). Peak matching for (M* + H), calcd for $C_{14}H_{16}O_6N_5$: 350.1100; found: 350.1110.

8-(3"-Furyl)guanosine (7d). This compound was obtained from 250 mg (0.69 mmol) of 6, 25.0 mL of HMDS, 2.5 mL of pyridine, 25 mg (0.03 mmol) of $PdCl_2(PPh_3)_2$ and 1.23 mg (3.45 mmol) of 3-tributylstannylfuran.³² After 40 h and HPLC purification (eluent 22:78) the yield was 140 mg (58%); mp 268 °C (dec). ¹H NMR (DMSO- d_6): δ 8.13 (dd, 1H, H2", J = 1.5, 0.9 Hz), 7.86 (dd, 1H, H5", J = 1.8, 1.5 Hz), 6.85 (dd, 1H, H4", J = 1.8, 0.9 Hz), 6.39 (s, NH₂), 5.71 (d, 1H, H1'), 4.98 (dd, 1H, H2'), 4.11 (dd, 1H, H3'), 3.86 (dd, 1H, H4'), 3.69 (dd, 1H, H5'). Peak matching for (M* + 1), calcd for $C_{14}H_{16}O_6N_3$: 350.1100; found: 350.1096.

Biochemistry

Inhibition of HIV-1, HSV-1 and influenza A multiplication were performed as XXT assays in MT4 cells (human T cell line), vero cells and MDCK cells (with Victoria 3/75 strain), respectively. Effect on cell growth was determined as an XTT assay on non-confluent HEL cells without presence of any virus. In the CMV assay, reduction in cytopathic effect caused by the virus was determined in MRC-5 cells (human embryonic cells). These assays were performed as previously described.³⁸ The CMV ELISA assay was an *in situ* determination of viral antigens in HEL cells and performed essentially as previously described.³⁹

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